UTILIZATION OF PTEROYLGLUTAMIC ACID CONJUGATES IN THE IN VITRO SYNTHESIS OF L. CITROVORUM ACTIVITY¹

The demonstration that liver slices convert synthetic pteroylglutamic acid (I) to a substance(s) possessing microbiological activity for L. citrovorum² led us to study the ability of chick liver tissue to convert pteroyldiglutamic acid (II) and pteroyltriglutamic acid (III) to a substance(s) possessing L. citrovorum activity. Using the procedure of Nichol and Welch² and employing the modifications reported previously³ it was observed (Table I) that liver slices from chicks deficient in I synthesized L. citrovorum activity, under the experimental conditions, at the same rate regardless of whether the substrate was I, II or III. The values presented were obtained using synthetic 5-formyl-5,6,7,8-tetrahydropteroylglutamic acid (leucovorin) as a standard. Ascorbic acid was added to the flasks (10 mg./flask) to augment the synthesis of L. citrovorum activity.²

TABLE I

L. Citrovorum Activity Synthesized by Pteroylglutamic Acid Deficient Chick Liver Tissue⁴

S11	bst	ta.	te

	т	TT	III			
None	$10\gamma/flask$	$\simeq 10 \gamma I / flask$	$\approx 10\gamma I/flask$			
0.12 ± 0.06	3.78 ± 0.51	3.50 ± 0.25	3.60 ± 0.34			
^a Values are the average of six experiments and are expressed as γ of leucovorin per g. liver slices, fresh wt., present after incubation. This amount of I is far in excess of that preded to extra the extern in corport to be the determined of the standard statement of the stateme						
needed to caturate the system in respect to I (The standard)						

needed to saturate the system in respect to I. The standard error is included.

Bioautographic analyses of the total L. citrovorum activity synthesized under these conditions were carried out. The modified procedure of Winsten and Eigen⁴ reported previously⁵ was employed. Under the experimental conditions two zones of growth were consistently observed. The faster moving spot had the same R_f value and could not be separated chromatographically from leucovorin. Both zones of growth were obtained regardless of whether the substrate was I, II or III.

These observations indicate that I, II and III apparently have a common metabolic pathway in the synthesis of L. *citrovorum* activity. It is furthermore indicated that more than one substrate possessing activity for L. *citrovorum* is synthesized by chick liver tissue.

DEPARTMENT OF BIOCHEMISTRY	L. S. DIETRICH
University of Wisconsin	W. J. Monson
Madison 6, Wisconsin	C. A. ELVEHJEM
RECEIVED MAY 16	1052

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(4) W. A. Winsten and E. Eigen, J. Biol. Chem., 184, 155 (1950).
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EVIDENCE FOR THE EXCHANGE OF HYDROXYL RADICAL WITH WATER¹

Sir:

Other work has shown that the reaction of H_2O_2 and O_3 produces a powerful le⁻-oxidizing agent which reacts with O_3 , H_2O_2 as well as with less reactive substances such as Cl⁻, Br⁻, HOAc, etc.² It has also been shown that the same species is generated by the reaction of HCOOH with O_3 .³ This species has been described as the hydroxyl radical, and the decomposition of O_3 which it catalyzes has been formulated^{2,4} as taking place by the steps

$$\begin{array}{c} HO + O_3 \longrightarrow HO_2 + O_2 \\ HO_2 + O_3 \longrightarrow HO + 2O_2 \end{array}$$

This reaction scheme suggests a simple tracer experiment for testing exchange of HO and H_2O , since the hydroxyl oxygen is converted to O_2 , which is known not to exchange readily with H_2O . Hydrogen peroxide chemistry offers no similar convenient means of studying the exchange, since the hydroxyl oxygen in reaction with H_2O_2 is presumably converted to water.

$$HO^* + H_2O_2 = H_2O^* + HO_2$$

This communication presents some results on the exchange of O_3 with H_2O induced by the reaction with H₂O₂. In all experiments ozonized oxygen of normal isotopic composition (N = mole fraction of $O^{18} = 2.000 \times 10^{-3}$; all isotopic compositions. quoted have been normalized to this value for N°) was left in contact with a liquid phase containing water enriched in O¹⁸ ($N = 14.6 \times 10^{-3}$). The ratio of gas volume to liquid was approximately 5. After a time, the gas was removed, dried, any residual O3 was decomposed and the isotopic composition determined. In an experiment on the direct exchange of O₃ with water, ozonized oxygen at atmospheric pressure, $7\% O_8$, was left in contact with enriched water, 0.04 M in HClO₄ for 5 days. The isotopic composition of the gas remained un-changed⁵ at 2.000×10^{-3} . In a typical experiment with H_2O_2 present, all conditions were the same except the liquid contained $9 \times 10^{-4} M H_2O_2$ (normal isotopic composition, $N = 1.997 \times 10^{-3}$). After 3 days, during which time 40% of the O₃ disappeared and 30% of the H₂O₂, the isotopic composition of the gas was found to be 2.050×10^{-3} .

The results quoted correspond to the exchange of about one-tenth of the oxygen contained in the O_3 which has decomposed. By the mechanism for decomposition which has been suggested, a maximum exchange of one-sixth of the O_3 oxygen can be expected.

It should be stressed that neither the earlier data nor the present data prove that the intermediate in question is HO. The present exchange data provide additional strong evidence however. H_2O_2 , O_2 and O_3 do not exchange at all rapidly with water.

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(2) H. Taube and W. C. Bray, THIS JOURNAL, 62, 3357 (1940).

(3) H. Taube, ibid., 63, 2453 (1941).

(4) J. Weiss, Trans. Faraday Soc., 31, 1547 (1934).

(5) In alkaline solution extensive exchange of osons and water does take place;

Sir: